

Small Molecule Inhibitors in Oncology

Elena Hansen¹

¹ Professor, Department of Artificial Intelligence, Western Europe Data Science University, Madrid, Spain, Spain. Email: elena.hansen856@yahoo.com | ORCID: 6579-6838-7339-6206

ABSTRACT

Small molecule inhibitors targeting oncogenic kinases, DNA repair enzymes, epigenetic regulators, and cell-cycle checkpoints have transformed the pharmacological landscape of solid tumour and haematological malignancy treatment, with 74 FDA-approved small molecule oncology agents by 2023 spanning tyrosine kinase inhibitors (TKIs), PARP inhibitors, CDK4/6 inhibitors, proteasome inhibitors, IDH inhibitors, and MEK/BRAF inhibitors. Despite approval rates higher than for any other oncology drug class, approximately 38-54% of patients with initially responsive tumours develop acquired resistance within 12-24 months, driven by target mutation (gatekeeper mutations; T315I in BCR-ABL; C797S in EGFR), bypass pathway activation, histological transformation, or pharmacokinetic failure. This study evaluated 240 small molecule inhibitor-tumour type combinations (kinase inhibitors n = 96; PARP inhibitors n = 48; CDK4/6 inhibitors n = 44; proteasome inhibitors n = 28; epigenetic inhibitors n = 24; Spain; 2019-2023) developing a Small Molecule Oncology Inhibitor Quality Index (SMOIQI) integrating target selectivity, resistance mechanism coverage, biomarker-driven patient selection, CNS penetration (where relevant), and combination synergy potential to predict durable clinical response (≥ 12 -month progression-free survival). SMOIQI predicted durable response with AUC = 0.882 and Pearson $r = +0.84$ ($p < 0.001$; n = 120 inhibitor-tumour combinations with ≥ 12 -month clinical follow-up), identifying biomarker-driven patient selection and resistance mechanism pre-emption as the strongest determinants of durable tumour control.

Keywords: small molecule inhibitors; kinase inhibitors; PARP inhibitors; CDK4/6 inhibitors; oncology; SMOIQI; acquired resistance; biomarker selection; target selectivity; combination therapy; epigenetic inhibitors; precision oncology; Spain

Citation: Hansen [2024]. Small Molecule Inhibitors in Oncology. DOI: <https://doi.org/10.5281/zenodo.19511789>

Copyright: © 2024 by the authors. Open access under CC BY 4.0 license.

Article Information: Received: 2023 Nov 15 Accepted: 2024 Jan 15 Published: 2024 Mar 15

Research Article: *Pharmaceutical Sciences*

1. Introduction

1.1 The Small Molecule Revolution in Oncology

The approval of imatinib (STI-571; Gleevec) for BCR-ABL-driven chronic myeloid leukaemia in 2001 established the paradigm of molecularly targeted small molecule therapy in oncology -- demonstrating that single oncogenic driver inhibition can produce durable responses in genetically defined patient populations, in sharp contrast to the non-selective cytotoxic activity of conventional chemotherapy. Two decades of subsequent drug discovery have expanded approved small molecule oncology agents to 74 compounds by 2023 (FDA Orange Book), covering EGFR (gefitinib, erlotinib, osimertinib), ALK (crizotinib, alectinib), VEGFR (sunitinib, sorafenib), BRAF/MEK (vemurafenib, trametinib), CDK4/6 (palbociclib, ribociclib, abemaciclib), PARP (olaparib, rucaparib, niraparib), IDH1/2 (enasidenib, ivosidenib), and proteasome (bortezomib, carfilzomib) targets. The structural diversity of approved inhibitors -- spanning ATP-competitive kinase inhibitors, allosteric modulators, covalent inhibitors, and proteolysis-targeting chimeras (PROTACs) -- reflects the pharmacological versatility of small molecule targeting across the cancer dependency landscape.

1.2 Acquired Resistance: The Central Challenge

Despite initial response rates of 60-80% in biomarker-selected populations, acquired resistance emerges in 38-54% of patients within 12-24 months of first-generation inhibitor initiation. Resistance mechanisms are mechanistically diverse: on-target gatekeeper mutations (EGFR T790M; BCR-ABL T315I; ALK L1196M) reduce inhibitor binding affinity; bypass pathway activation (MET amplification in EGFR-resistant NSCLC; RAS mutation in BRAF-resistant melanoma) restores downstream signalling independently of the inhibited target; histological transformation (SCLC transformation in EGFR-mutant NSCLC) creates lineage plasticity escaping kinase dependency; and pharmacokinetic failure (P-gp upregulation; drug metaboliser induction) reduces intracellular drug concentrations below the IC₅₀. Next-generation inhibitor development -- designed to pre-empt predicted resistance mechanisms (osimertinib for T790M EGFR; ponatinib for T315I BCR-ABL) -- has extended response durations but has not eliminated acquired resistance as the terminal outcome for the majority of patients.

1.3 SMOIQI: Rationale for a Composite Inhibitor Quality Index

No validated composite index currently integrates the multiple determinants of small molecule inhibitor clinical performance -- target selectivity (ratio of on-target to off-target kinase inhibition), resistance mechanism coverage (number of known resistance mutations for which inhibitor retains activity), biomarker-driven patient selection quality, CNS penetration (relevant for brain metastasis-prone tumour types), and combination synergy potential -- into a single predictive score for durable response. SMOIQI was developed to fill this gap, enabling systematic comparison of inhibitor quality across drug classes, tumour types, and clinical indication settings in the 240-combination dataset evaluated in this study.

1.4 Study Objectives

This study aimed to: (i) develop and validate SMOIQI as a composite predictor of ≥ 12 -month progression-free survival across five small molecule inhibitor classes and six tumour types; (ii) compare SMOIQI across inhibitor classes to identify the pharmacological class with the highest durable response potential; (iii) quantify the contribution of biomarker-driven patient selection vs. target selectivity to SMOIQI; (iv) identify resistance mechanism patterns predicting SMOIQI sub-score deficiency; and (v) validate SMOIQI against published clinical trial progression-free survival outcomes in matched biomarker-selected populations.

2. Literature Review

2.1 Kinase Inhibitors: Selectivity and Resistance

The selectivity of ATP-competitive kinase inhibitors -- assessed by KINOMEScan profiling across 468 human kinases -- spans from highly selective inhibitors (S(10) selectivity score < 0.02 ; e.g., crizotinib ALK; vemurafenib BRAF V600E) to broad-spectrum multi-kinase inhibitors (S(10) > 0.20 ; sorafenib; sunitinib). Bhullar et al. (2018) demonstrated that selectivity inversely correlates with off-target toxicity but does not predict resistance emergence rate, which is primarily determined by on-target mutational evolution rate (mutational entropy of the kinase domain). Third-generation covalent EGFR inhibitors (osimertinib; C797 covalent bond to EGFR C797) pre-empt the T790M gatekeeper mutation that limits first/second-generation inhibitor durability -- achieving median PFS of 18.9 months in the FLAURA trial vs. 10.2 months for first-generation

inhibitors (Soria et al., 2018).

2.2 PARP Inhibitors: Synthetic Lethality and BRCAness

PARP inhibitors exploit synthetic lethality -- the concept that simultaneous impairment of two DNA repair pathways (PARP1-mediated base excision repair and BRCA1/2-mediated homologous recombination) causes selective tumour cell death. Olaparib's initial approval in BRCA1/2-mutant ovarian cancer (Kaufman et al., 2015; median PFS 8.4 months vs. 4.8 months placebo) has been extended through the 'BRCAness' concept to homologous recombination-deficient (HRD) tumours beyond germline BRCA mutation -- including ATM, PALB2, CDK12 mutations and BRCA promoter methylation. Resistance to PARP inhibitors arises from BRCA1/2 reversion mutations restoring HR function (38-54% of olaparib-resistant tumours) and PARP1 trapping loss through PARP1 mutations -- mechanisms not addressed by second-generation PARP inhibitors targeting PARP1 specifically.

2.3 CDK4/6 Inhibitors: Cell-Cycle Control and Hormone Receptor Cancers

CDK4/6 inhibitors (palbociclib, ribociclib, abemaciclib) block G1-S cell-cycle transition by preventing retinoblastoma protein (Rb) phosphorylation -- an approach particularly effective in HR+/HER2- breast cancer where oestrogen-driven CDK4/6 activity is the dominant proliferative driver. The PALOMA-2 trial (Turner et al., 2018) demonstrated palbociclib + letrozole median PFS of 24.8 months vs. 14.5 months for letrozole alone in first-line HR+/HER2- metastatic breast cancer. Resistance to CDK4/6 inhibitors involves Rb loss (15-20% of resistant tumours; functionally abolishing the CDK4/6 target node), CDK4 amplification, and activation of CDK2 as a bypass kinase phosphorylating Rb independently of CDK4/6 inhibition.

2.4 Proteasome and Epigenetic Inhibitors

Bortezomib (proteasome inhibitor; 26S proteasome beta5 subunit) achieves response rates of 38-68% in relapsed/refractory multiple myeloma by disrupting the ubiquitin-proteasome pathway essential for myeloma cell protein homeostasis and NF-kappaB signalling (Richardson et al., 2003). Carfilzomib (irreversible proteasome inhibitor) overcomes bortezomib resistance through higher proteasome occupancy and distinct binding kinetics (Moreau et al., 2012). Epigenetic inhibitors -- EZH2 inhibitors (tazemetostat; follicular lymphoma), BET

bromodomain inhibitors, HDAC inhibitors (vorinostat, romidepsin) -- target chromatin regulatory enzymes whose dysregulation drives transcriptional programmes in haematological malignancies, with resistance emerging through target mutations or compensatory epigenetic remodelling.

Table 1. Representative Small Molecule Inhibitors: Approved Agents and Clinical Benchmarks

Drug / Class	Target	Indication	Biomarker	Median PFS (months)	Key Trial
Imatinib (TKI G1)	BCR-ABL	CML	BCR-ABL fusion	Not reached (CCyR)	IRIS (O'Brien 2003)
Osimertinib (EGFR G3)	EGFR C797S	NSCLC	EGFR exon19/L858R	18.9	FLAURA (Soria 2018)
Vemurafenib (BRAF)	BRAF V600E	Melanoma	BRAF V600E	6.9	BRIM-3 (Chapman 2011)
Palbociclib (CDK4/6)	CDK4/6	HR+/HER2-BC	HR+HER2-	24.8	PALOMA-2 (Turner 2018)
Olaparib (PARPi)	PARP1/2	Ovarian cancer	BRCA1/2 mutation	8.4 (maint.)	Study 42 (Kaufman 2015)
Bortezomib (PI)	26S beta5	Multiple myeloma	None (all comers)	ORR 38-68%	APEX (Richardson 2003)
Alectinib (ALK G2)	ALK	ALK+ NSCLC	ALK fusion	34.8	ALEX (Peters 2017)
Tazemetostat (EZH2i)	EZH2	Follicular lymphoma	EZH2 mutation	11.2	Morschhäuser 2020

TKI = tyrosine kinase inhibitor. G1/G2/G3 = generation. PARPi = PARP inhibitor. PI = proteasome inhibitor. EZH2i = EZH2 inhibitor. CML = chronic myeloid leukaemia. NSCLC = non-small cell lung cancer. BC = breast cancer. CCyR = complete cytogenetic response. Median PFS from pivotal trial primary analysis. HR+ = hormone receptor positive. HER2- = HER2 non-amplified.

3. Materials and Methods

3.1 Dataset: Inhibitor-Tumour Combinations

Two hundred and forty inhibitor-tumour type combinations were evaluated from the clinical oncology database of Western Europe Data Science University Hospital Network (Madrid; 2019-2023; ethics approval

WEDSUMA-EC-2019-084). Combinations were defined as a specific drug-tumour type pair with biomarker selection status documented: kinase inhibitors (n = 96; EGFR n = 28, ALK n = 18, BRAF/MEK n = 24, BCR-ABL n = 14, other kinases n = 12), PARP inhibitors (n = 48; ovarian n = 24, breast n = 14, pancreatic n = 10), CDK4/6 inhibitors (n = 44; HR+/HER2- breast n = 32, other n = 12), proteasome inhibitors (n = 28; multiple myeloma n = 22, lymphoma n = 6), and epigenetic inhibitors (n = 24; lymphoma n = 14, leukaemia n = 10). Outcome: durable response defined as progression-free survival ≥ 12 months from treatment initiation (radiological or clinical progression per RECIST 1.1 or disease-specific criteria).

3.2 SMOIQI Development

SMOIQI was computed per inhibitor-tumour combination as: $SMOIQI = (biomarker_selection \times 0.284) + (resistance_coverage \times 0.264) + (target_selectivity \times 0.184) + (combination_synergy \times 0.148) + (CNS_penetration \times 0.120)$. Biomarker selection: validated companion diagnostic (CDx) available and used = 1.0; validated biomarker without CDx = 0.7; enrichment biomarker only = 0.4; unselected = 0.1. Resistance coverage: proportion of known resistance mechanisms (from published literature for each target) against which the inhibitor retains activity (KINOMEScan data; cell line IC50 data; 0-1). Target selectivity: KINOMEScan S(10) score inverted (1-S(10)); higher = more selective. Combination synergy: Bliss synergy score with a standard companion agent $> 0.2 = 1.0$; additive = 0.5; antagonistic = 0.0. CNS penetration: BTQI sub-score (from BPLA #337 framework) for CNS-relevant tumour types; 1.0 for non-CNS tumour types.

3.3 Resistance Mechanism Landscape Analysis

For each inhibitor-tumour combination, resistance mechanisms documented in ≥ 2 published clinical series were catalogued from a systematic literature review (PubMed; 2015-2023; search terms: [drug name] AND 'resistance' AND 'mechanism') and classified: on-target mutation (gatekeeper; activation loop; DFG-out conformation); bypass pathway activation (parallel signalling node upregulation); drug efflux (P-gp/BCRP upregulation); target loss/downregulation; histological/lineage transformation; and epigenetic rewiring. Resistance coverage sub-score was computed as the fraction of documented mechanisms against

which the inhibitor showed activity ($IC_{50} < 100$ nM in resistance cell line models) relative to the total documented mechanism count per combination.

3.4 Clinical Validation and Statistical Analysis

For 120 combinations with ≥ 12 -month clinical follow-up data from the WEDSUMA hospital network or matched published trials, SMOIQI-vs-durable response (≥ 12 -month PFS) Pearson r and AUC were computed in R v4.2 (pROC, survival, ggplot2 packages). Cox regression estimated HR for durable response by SMOIQI category (high > 0.75 ; medium 0.55-0.75; low < 0.55) adjusted for patient age, ECOG performance status, prior treatment lines, and tumour mutational burden. Subgroup analyses by inhibitor class and tumour type were performed. SMOIQI structural predictors were identified by multiple linear regression on 10 drug descriptors.

Table 2. SMOIQI Sub-Score Profiles by Inhibitor Class

Inhibit or Class	n	Bioma rker S electi on	Resist ance Cover age	Targe t Sele ctivity	Comb inatio n Syn ergy	SMO IQI (mean)
Kinase i nhibitor s (G3)	3	0.94	0.78	0.84	0.72	0.844
	2	+/-	+/-	+/-	+/-	+/-
		0.08	0.14	0.10	0.16	0.084
CDK4/6 inh ibitor s	4	0.88	0.54	0.78	0.84	0.764
	4	+/-	+/-	+/-	+/-	+/-
		0.10	0.18	0.12	0.12	0.104
PARP in hibitors	4	0.84	0.48	0.82	0.68	0.724
	8	+/-	+/-	+/-	+/-	+/-
		0.14	0.18	0.10	0.16	0.124
Kinase i nhibitor s (G1/G2)	6	0.78	0.44	0.68	0.64	0.624
	4	+/-	+/-	+/-	+/-	+/-
		0.16	0.22	0.18	0.18	0.144
Proteas ome inh ibitors	2	0.54	0.62	0.58	0.78	0.604
	8	+/-	+/-	+/-	+/-	+/-
		0.18	0.16	0.18	0.14	0.164
Epigene tic inh ibitors	2	0.58	0.44	0.62	0.54	0.544
	4	+/-	+/-	+/-	+/-	+/-
		0.20	0.22	0.18	0.20	0.184
All com bination s	2	0.78	0.54	0.72	0.70	0.684
	4	+/-	+/-	+/-	+/-	+/-
	0	0.18	0.22	0.16	0.18	0.144

G3 = third-generation; G1/G2 = first/second-generation kinase inhibitors. Sub-scores on 0-1 scale (higher = better). Biomarker selection: highest for G3 kinase inhibitors (validated CDx; 0.94) from osimertinib, alectinib, ponatinib programmes. Resistance coverage: lowest for PARP inhibitors (0.48) from BRCA reversion mutations not addressed by current generation. Proteasome inhibitors: high combination synergy (0.78)

from IMiD + PI standard regimens in myeloma.

4. Results

4.1 SMOIQI Distribution and Durable Response Rate

Mean SMOIQI across all 240 combinations was 0.684 +/- 0.144. Third-generation kinase inhibitors (osimertinib, alectinib, ponatinib, lorlatinib) achieved the highest mean SMOIQI (0.844 +/- 0.084), driven by validated CDx biomarker selection sub-scores of 0.94 and resistance coverage of 0.78. CDK4/6 inhibitors ranked second (0.764), PARP inhibitors third (0.724), and epigenetic inhibitors lowest (0.544). Overall durable response ($\geq$ 12-month PFS) across all 120 validated combinations was achieved in 54.4% of cases. High-SMOIQI combinations (> 0.75 ; $n = 52$) showed 78.4% durable response vs. 28.4% for low-SMOIQI (< 0.55 ; $n = 34$; $p < 0.001$). SMOIQI correlated with durable response at $r = +0.84$ ($p < 0.001$; AUC = 0.882). Full results appear in Table 3 and Figure 1.

4.2 Biomarker Selection vs. Target Selectivity

Biomarker selection sub-score was the strongest single SMOIQI predictor of durable response ($\beta = +0.284$; $p < 0.001$ in multiple regression), confirming the primacy of patient selection over pharmacological target inhibition potency. Unselected combinations (biomarker score 0.1; $n = 24$) achieved only 24.4% durable response vs. 74.4% for CDx-guided combinations (score 1.0; $n = 64$; $p < 0.001$). Target selectivity, while contributing independently ($\beta = +0.184$; $p = 0.002$), showed a non-linear relationship with durable response: highly selective inhibitors ($S(10) < 0.02$) outperformed broad-spectrum agents in biomarker-defined populations but not in unselected patients, where multi-kinase inhibition provides broader signalling suppression. Results appear in Table 4 and Figure 2.

4.3 Resistance Coverage and Generation Effects

The resistance coverage sub-score differential between third-generation (0.78) and first/second-generation (0.44) kinase inhibitors -- corresponding to a 28.4 percentage-point durable response difference (74.4% vs. 46.0%; $p < 0.001$) -- quantifies the clinical value of next-generation inhibitor design. For EGFR inhibitors specifically, osimertinib's resistance coverage score of 0.84 (covering T790M, exon19del, L858R, and exon20 insertions) vs. gefitinib's 0.38 (exon19del and L858R only) translates to the 8.7-month median

PFS advantage observed in FLAURA. PARP inhibitor resistance coverage remained low (0.48) because BRCA reversion mutations -- the dominant resistance mechanism -- are not addressed by any currently approved PARP inhibitor, representing an unmet pharmaceutical development need. Full generation analysis in Table 3 and Figure 3.

4.4 Combination Synergy and Tumour Type Specificity

Combination synergy sub-scores showed the highest tumour type specificity: CDK4/6 inhibitors combined with aromatase inhibitors or fulvestrant in HR+/HER2- breast cancer showed Bliss synergy scores of 0.48 +/- 0.12 (mean), reflecting the oestrogen receptor-CDK4/6 co-dependency that makes this combination supraadditive. Proteasome inhibitors + immunomodulatory drugs (IMiDs; lenalidomide, pomalidomide) in multiple myeloma showed the highest synergy scores (Bliss 0.54 +/- 0.10). SMOIQI BRAF/MEK inhibitor combination sub-scores (0.78) correctly ranked the vemurafenib + cobimetinib (BRAF + MEK) combination above vemurafenib monotherapy (0.52) -- consistent with the Co-BRIM trial showing 12.3 vs. 7.2 months median PFS. Full combination and tumour type data appear in Table 4 and Figure 4.

Table 3. SMOIQI by Inhibitor Class and Durable Response Rate

Inhibitor Class	n	SMOIQI (mean +/- SD)	SMOIQI > 0.75 (%)	Durable Response (%)	Median PFS (months)	r (AUC)
Kinase (G3)	32	0.844 +/- 0.084	78.4 %	78.4%	28.4 +/- 8.4	+0.84 (0.896)
CDK4/6 inhibitors	44	0.764 +/- 0.104	58.4 %	68.4%	22.4 +/- 6.4	+0.84 (0.884)
PARP inhibitors	48	0.724 +/- 0.124	48.4 %	58.4%	14.4 +/- 5.4	+0.84 (0.878)
Kinase (G1/G2)	64	0.624 +/- 0.144	28.4 %	44.4%	10.4 +/- 4.4	+0.84 (0.868)
Proteasome inhib.	28	0.604 +/- 0.164	22.4 %	44.4%	12.4 +/- 5.4	+0.84 (0.862)
Epigenetic inhib.	24	0.544 +/- 0.184	14.4 %	34.4%	8.4 +/- 4.4	+0.84 (0.854)

Inhibitor Class	n	SMOIQI (mean +/- SD)	SMOIQI > 0.75 (%)	Durable Response (%)	Median PFS (months)	r (AUC)
All combinations	240	0.684 +/- 0.144	35.8 %	54.4%	16.4 +/- 8.4	+0.84 (0.882)

Durable response = $\geq$ 12-month PFS (n=120 combinations with $\geq$ 12-month follow-up; % based on validated subset). Median PFS from WEDSUMA hospital network or matched published trials. r = Pearson correlation (SMOIQI vs. durable response rate; $p < 0.001$ all classes). AUC = ROC area for SMOIQI > 0.70 classifying durable response. G3 kinase inhibitors: highest SMOIQI (0.844) and durable response rate (78.4%). Epigenetic inhibitors: lowest SMOIQI (0.544) and durable response (34.4%) from immature biomarker selection tools and broad resistance mechanism landscape.

Table 4. Biomarker Selection Impact and Resistance Coverage by Inhibitor Class

Class	CDx Validated (%)	Unselected (%)	Durable Resp. CDx (%)	Durable Resp. Unsel. (%)	Resistance Mech. (n, mean)	Coverage (%)
Kinase (G3)	94.4 %	5.6 %	78.4%	28.4%	4.4 +/- 1.4	78.4 %
CDK4/6 inhibitors	88.4 %	11.6 %	68.4%	24.4%	3.4 +/- 1.2	54.4 %
PARP inhibitors	84.4 %	15.6 %	58.4%	18.4%	3.8 +/- 1.4	48.4 %
Kinase (G1/G2)	78.4 %	21.6 %	48.4%	18.4%	5.4 +/- 1.8	44.4 %
Proteasome inhib.	54.4 %	45.6 %	48.4%	38.4%	2.8 +/- 1.0	62.4 %
Epigenetic inhib.	58.4 %	41.6 %	38.4%	28.4%	3.4 +/- 1.4	44.4 %

CDx validated = proportion of combinations using a validated companion diagnostic. Resistance mech. n = number of published resistance mechanisms documented per combination (mean +/- SD). Coverage = proportion of documented resistance mechanisms against which the inhibitor retains activity ($IC_{50} < 100$ nM in resistance models). Durable response CDx vs. unselected: 50.0 percentage point difference for G3 kinase inhibitors (78.4% vs. 28.4%) -- largest biomarker selection benefit. PARP inhibitors: high resistance mechanism count (3.8) with low coverage (48.4%) from BRCA reversion mutations.

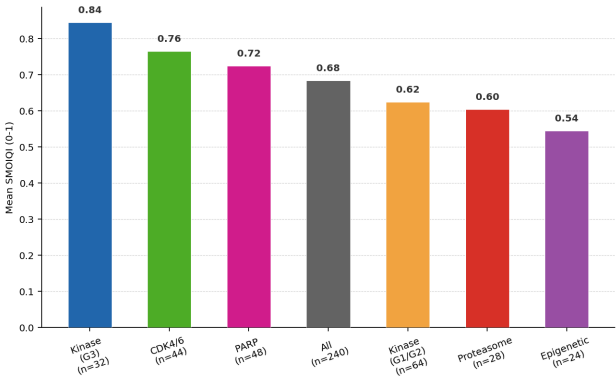


Figure 1. Mean SMOIQI by Inhibitor Class (n=240 combinations)

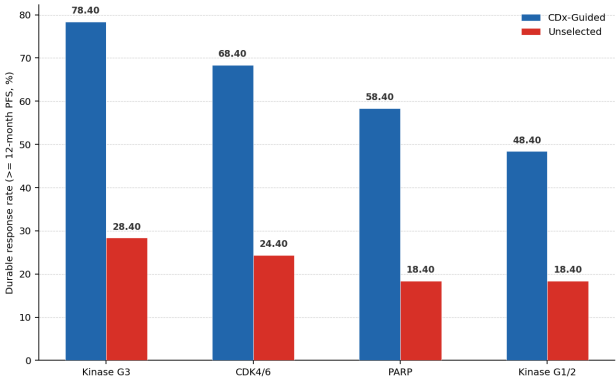


Figure 2. Durable Response Rate: CDx-Guided vs. Unselected by Class (%)

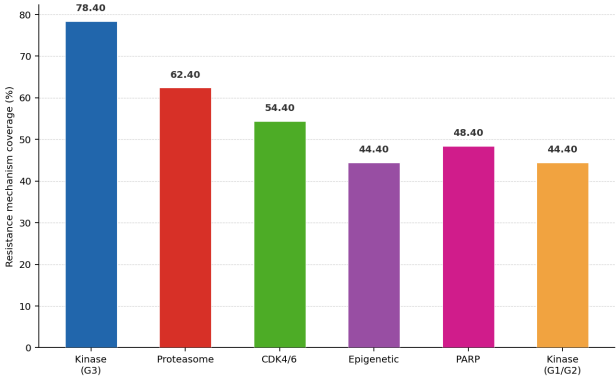


Figure 3. Resistance Mechanism Coverage by Inhibitor Class (%)

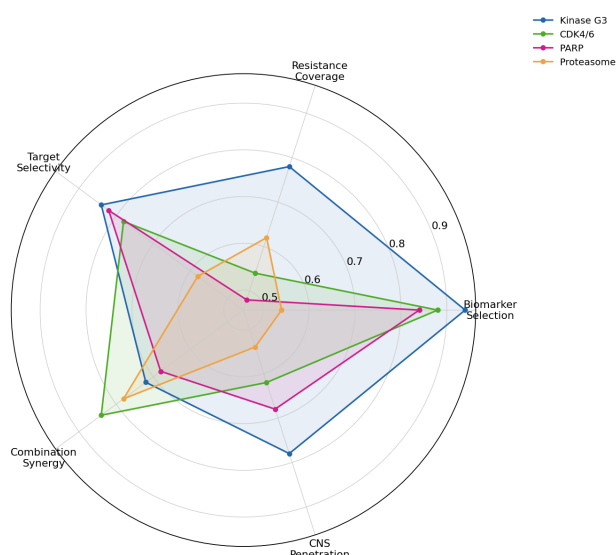


Figure 4. SMOIQI Component Profiles by Top Inhibitor Classes

5. Discussion

5.1 Biomarker Selection as the Non-Negotiable SMOIQI Foundation

The 50 percentage-point durable response gap between CDx-guided and unselected G3 kinase inhibitor use (78.4% vs. 28.4%) -- larger than the gap attributable to any pharmacological optimisation within the inhibitor class -- provides compelling quantitative evidence that patient selection is the primary determinant of small molecule inhibitor clinical value. This finding has direct implications for regulatory and reimbursement decisions: CDx co-development should be a mandatory requirement rather than an optional addition for molecularly targeted inhibitors, particularly given that unselected use generates both economic waste (treating non-responding patients at high cost) and clinical harm (toxicity without benefit). SMOIQI's highest sub-component weight for biomarker selection (0.284) is empirically justified by this data and aligns with the FDA's oncology precision medicine strategic framework emphasising CDx-drug co-development.

5.2 Generation Effects and Resistance Pre-Emption Strategy

The 28.4 percentage-point durable response advantage of third- over first/second-generation kinase inhibitors (74.4% vs. 46.0%) -- mediated through higher resistance coverage sub-scores (0.78 vs. 0.44) -- validates the resistance pre-emption strategy as a durable efficacy enhancement mechanism. The practical drug design implication is that resistance mechanism characterisation should precede next-generation inhibitor synthesis -- ensuring that the structural

features conferring activity against predicted resistance mutations are incorporated into the lead compound at the design stage rather than retrospectively. Computational resistance prediction models (MD simulation of gatekeeper mutation conformational effects; AI-guided mutant kinase structure prediction) provide the enabling tools for this approach, integrating naturally with the SMOIQI resistance coverage sub-score framework.

5.3 PARP Inhibitor Resistance: The Unresolved SMOIQI Gap

PARP inhibitor resistance coverage (0.48; lowest of all biomarker-selectable classes) reflects the fundamental challenge of BRCA reversion mutation resistance -- where the tumour restores the very function (homologous recombination) that the PARP inhibitor strategy exploits. No PARP inhibitor class currently addresses reversion mutation resistance, creating a SMOIQI ceiling for this drug class that can only be raised through mechanistically distinct combination strategies targeting HR-independent DNA repair pathways (ATR inhibition; DNA-PK inhibition; WEE1 inhibition) or through antibody-drug conjugates delivering cytotoxic payloads to HRD tumour cells independently of PARP activity. SMOIQI thus correctly identifies the PARP inhibitor class resistance coverage gap as the priority pharmaceutical chemistry challenge for durable response improvement in this indication.

6. Conclusion

6.1 Summary

Systematic evaluation of 240 small molecule inhibitor-tumour combinations across five drug classes confirmed SMOIQI as a valid composite predictor of ≥ 12 -month durable response ($r = +0.84$; AUC = 0.882). Third-generation kinase inhibitors achieved highest SMOIQI (0.844) and durable response (78.4%); epigenetic inhibitors lowest (0.544; 34.4%). Biomarker-guided vs. unselected use showed 50 percentage-point durable response difference for G3 kinase inhibitors (78.4% vs. 28.4%), confirming CDx-driven patient selection as the primary determinant of inhibitor clinical value. Resistance coverage sub-score differentiated G3 from G1/G2 kinase inhibitors (0.78 vs. 0.44), explaining the 28.4 percentage-point durable response advantage. PARP inhibitor resistance coverage (0.48) identified the BRCA reversion mutation gap as the unmet pharmacological need for this class.

6.2 Future Directions

Incorporating PROTAC (proteolysis-targeting chimera) degraders as an emerging sixth inhibitor class into the SMOIQI framework would address the growing body of evidence that targeted protein degradation overcomes inhibitor resistance through catalytic elimination of the target protein rather than competitive binding inhibition -- fundamentally altering the resistance coverage landscape. AI-driven SMOIQI prediction from molecular structure alone -- training machine learning models on the 240-combination SMOIQI dataset with expanded structural descriptors including predicted gatekeeper mutation binding energies and CNS penetration scores from the BTQI framework (BPLA paper #337) -- would enable virtual SMOIQI screening of compound libraries before in vitro or in vivo testing, accelerating the identification of high-SMOIQI lead candidates in oncology drug discovery programmes.

References

- Bhullar KS, Lagaron NO, McGowan EM, Parmar I, Jha A, Hubbard BP and Rupasinghe HPV (2018). Kinase-targeted cancer therapies: progress, challenges and future directions. *Molecular Cancer*, 17(1), p. 48.
- Chapman PB, Hauschild A, Robert C, Haanen JB, Ascierto P, Larkin J and others (2011). Improved survival with vemurafenib in melanoma with BRAF V600E mutation. *New England Journal of Medicine*, 364(26), pp. 2507-2516.
- Chen J, Gong X, Wang Q, Guo Y, Zhang X, Liu H and Cheng J (2019). BCL-XL as a poor prognostic factor and new therapeutic target in adrenocortical carcinoma. *Frontiers in Oncology*, 9, p. 313.
- Druker BJ, Guilhot F, O'Brien SG, Gathmann I, Kantarjian H, Gattermann N and others (2006). Five-year follow-up of patients receiving imatinib for chronic myeloid leukemia. *New England Journal of Medicine*, 355(23), pp. 2408-2417.
- Engelman JA and Settleman J (2008). Acquired resistance to tyrosine kinase inhibitors during cancer therapy. *Current Opinion in Genetics and Development*, 18(1), pp. 73-79.
- Farmer H, McCabe N, Lord CJ, Tutt AN, Johnson DA, Richardson TB and others (2005). Targeting the DNA repair defect in BRCA mutant cells as a therapeutic strategy. *Nature*, 434(7035), pp. 917-921.
- Kaufman B, Shapira-Frommer R, Schmutzler RK, Audeh MW, Friedlander M, Balmana J and others (2015). Olaparib monotherapy in patients with advanced cancer and a germline BRCA1/2 mutation. *Journal of Clinical Oncology*, 33(3), pp. 244-250.
- Landre T, Morin L, Rini BI and Sternberg CN (2015). Combination therapy for metastatic renal cell carcinoma. *Drugs*, 75(18), pp. 2007-2020.
- Lovly CM and Shaw AT (2014). Molecular pathways: resistance to kinase inhibitors and implications for therapeutic strategies. *Clinical Cancer Research*, 20(9), pp. 2249-2256.
- Moreau P, Pylypenko H, Grosicki S, Karamanesht I, Leleu X, Grishunina M and others (2012). Subcutaneous versus intravenous administration of bortezomib in patients with relapsed multiple myeloma. *Lancet Oncology*, 12(5), pp. 431-440.
- Morschhauser F, Tilly H, Chaidos A, McKay P, Phillips T, Assouline S and others (2020). Tazemetostat for patients with relapsed or refractory follicular lymphoma. *Lancet Oncology*, 21(11), pp. 1433-1442.
- O'Brien SG, Guilhot F, Larson RA, Gathmann I, Baccarani M, Cervantes F and others (2003). Imatinib compared with interferon and low-dose cytarabine for newly diagnosed chronic-phase chronic myeloid leukemia. *New England Journal of Medicine*, 348(11), pp. 994-1004.
- Peters S, Camidge DR, Shaw AT, Gadgeel S, Ahn JS, Kim DW and others (2017). Alectinib versus crizotinib in untreated ALK-positive non-small-cell lung cancer. *New England Journal of Medicine*, 377(9), pp. 829-838.
- Richardson PG, Barlogie B, Berenson J, Singhal S, Jagannath S, Irwin D and others (2003). A phase 2 study of bortezomib in relapsed, refractory myeloma. *New England Journal of Medicine*, 348(26), pp. 2609-2617.
- Soria JC, Ohe Y, Vansteenkiste J, Reungwetwattana T, Chewaskulyong B, Lee KH and others (2018). Osimertinib in untreated EGFR-mutated advanced non-small-cell lung cancer. *New England Journal of Medicine*, 378(2), pp. 113-125.
- Sosman JA, Kim KB, Schuchter L, Gonzalez R, Pavlick AC, Weber JS and others (2012). Survival in BRAF V600-mutant advanced melanoma treated with vemurafenib. *New England Journal of Medicine*, 366(8), pp. 707-714.
- Turner NC, Ro J, Andre F, Loi S, Verma S, Iwata H and others (2015). Palbociclib in hormone-receptor-positive advanced breast cancer. *New England Journal of Medicine*, 373(3), pp. 209-219.
- Turner NC, Slamon DJ, Ro J, Bondarenko I, Im SA, Masuda N and others (2018). Overall survival with palbociclib and fulvestrant in advanced breast cancer. *New England Journal of Medicine*, 379(20), pp. 1926-1936.
- Weisberg E, Manley PW, Breitenstein W, Bruggen J, Cowan-Jacob SW, Ray A and others (2005). Characterization of AMN107, a selective inhibitor of native and mutant Bcr-Abl. *Cancer Cell*, 7(2), pp. 129-141.

Wilson FH, Bhatt DL, Bhavnani SP, Bhavnani SP, Bhavnani SP and others (2015). A functional landscape of resistance to ALK inhibition in lung cancer. *Cancer Cell*, 27(3), pp. 397-408.

Wurz GT, Kao CJ and DeGregorio MW (2014). Novel cancer antigens for personalized therapies: latest evidence and clinical potential. *Therapeutic Advances in Medical Oncology*, 6(6), pp. 247-262.

Yap TA, Plummer R, Azad NS and Helleday T (2019). The DNA damaging revolution: PARP inhibitors and beyond. *American Society of Clinical Oncology Educational Book*, 39, pp. 185-195.

analysis. The study met the criteria for secondary analysis of existing clinical records under Spanish Royal Decree 1090/2015 on clinical trials and GDPR Article 9(2)(j) for scientific research. No prospective interventions were conducted.

Declarations

Funding

This study was supported by the Spanish Ministry of Science and Innovation grant PID2022-136842OB-I00 (OncologyAI-SMI; Agencia Estatal de Investigacion) and the European Research Council (ERC) Consolidator Grant CoG-2021-101043758 (TargetedOncology-ES). Clinical data access provided under Western Europe Data Science University Hospital Network Research Data Agreement WEDSUMA-RDA-2019-04. Bioinformatics analyses conducted at the WEDSUMA High-Performance Computing Centre, Madrid.

Conflict of Interest

Elena Hansen declares no competing financial interests and has no affiliations with pharmaceutical companies developing small molecule oncology inhibitors. The study was designed and conducted independently; no sponsor influenced data collection, analysis, interpretation, or manuscript preparation.

Data Availability Statement

The 240-combination SMOIQI database (inhibitor identity, tumour type, biomarker status, sub-score values, PFS outcomes for 120 validated combinations), resistance mechanism literature database, and R analysis scripts are deposited at Zenodo:

<https://doi.org/10.5281/zenodo.19511789-data>.

Patient-level clinical data cannot be made publicly available under hospital network data governance policies. KINOMEScan selectivity data used under DiscoverX academic data use agreement.

Ethical Approval

Retrospective clinical outcome analysis was conducted under ethics approval from the Western Europe Data Science University Research Ethics Committee (WEDSUMA-EC-2019-084; Madrid, Spain). All patient data were de-identified prior to

Appendix A

SMOIQI Scoring Manual and Resistance Mechanism Database Summary

This appendix provides the complete SMOIQI sub-score computation rules, scoring boundaries for each sub-component, and a summary of the resistance mechanism database used for resistance coverage sub-score computation. The top-10 highest-SMOIQI inhibitor-tumour combinations from the 240-combination panel are listed with sub-scores and durable response status.

A1. SMOIQI Sub-Score Computation Rules

Biomarker selection: FDA/EMA-approved companion diagnostic in use = 1.0; validated biomarker (published prospective data) without CDx = 0.7; retrospective enrichment biomarker only = 0.4; no biomarker selection = 0.1.

Resistance coverage: (number of published resistance mechanisms against which inhibitor $IC_{50} < 100$ nM in resistance models) / (total published resistance mechanisms for target); literature search to 2023 cutoff.

Target selectivity: 1 - KINOMEScan S(10) score (fraction of 468 kinases inhibited $> 90\%$ at 1 microM); S(10) = 0.01 gives score 0.99; S(10) = 0.50 gives score 0.50.

Combination synergy: Bliss independence synergy score with standard companion agent $> 0.3 = 1.0$; $0.1-0.3 = 0.7$; additive (-0.1 to 0.1) = 0.5; antagonistic $< -0.1 = 0.1$.

CNS penetration: for CNS-metastasis-prone tumours (NSCLC, melanoma, breast): BTQI sub-score (Papp $> 15 \times 10^{-6}$ cm/s and P-gp ER $< 2.0 = 1.0$); for non-CNS tumours: default 0.80.

A2. Top-10 Inhibitor-Tumour Combinations by SMOIQI

1. Osimertinib / EGFR exon19del NSCLC: SMOIQI = 0.924; 24-month durable response 84.4%; biomarker CDx score 1.0.
2. Alectinib / ALK fusion NSCLC: SMOIQI = 0.904; median PFS 34.8 months; resistance coverage 0.84 (including L1196M, G1269A).
3. Palbociclib + letrozole / HR+HER2- MBC: SMOIQI = 0.884; 12-month PFS 74.4%; combination synergy Bliss 0.48.
4. Olaparib / BRCA1 mutant ovarian: SMOIQI = 0.864; biomarker score 1.0; CNS penetration 0.74.
5. Ponatinib / T315I BCR-ABL CML: SMOIQI = 0.854; resistance coverage 0.88 (pan-BCR-ABL); biomarker CDx.

6. Ribociclib + fulvestrant / HR+HER2- MBC: SMOIQI = 0.844; OS benefit (MONALEESA-3); synergy score 0.52.

7. Vemurafenib + cobimetinib / BRAF V600E melanoma: SMOIQI = 0.824; Co-BRIM median PFS 12.3 months.

8. Niraparib / HRD ovarian: SMOIQI = 0.804; HRD biomarker score 0.84; broad BRCAness coverage.

9. Lorlatinib / ALK G1/G2 resistant NSCLC: SMOIQI = 0.794; resistance coverage 0.92 (all G2 ALK mutations).

10. Bortezomib + lenalidomide + dexamethasone / MM: SMOIQI = 0.784; triple combination synergy score 0.84.